PATHOLOGICAL, MALE REPRODUCTIVITY AND RESIDUES OF DIMETHOATE TOXICITY IN ALBINO RATS

Abou Salem, M. E. *; El-Mashad, A. I. ** and Moustafa, S. A. **

* Department of Animal medicine (Forensic Med. & Toxicology)
** Department of Pathology Faculty of Vet. Medicine, Zagazig University, Benha branch.

ABSTRACT

The current study explores the effect of the organophosphorus compound (dimethoate) on male reproductive efficiency, tissue residues and its pathological picture. Dimethoate was given either directly to albino rats at doses of 1/10, 1/20, 1/40 of LD50 and also given green sprayed forage along 65 consecutive days. The results indicated that dimethoate altered the biochemical parameters of serum indicating liver and kidney dysfunction. Also semen picture and the weight of male sexual organs (testes, prostate and seminal vesicle) together with lower testosterone level gave indication for lower reproductive efficiency. Tissue dimethoate residues were detected in liver, testes and skeletal muscles. Concerning to pathological findings dimethoate induced degenerative changes in liver, kidneys and heart in the form of cloudy swelling, vacuolar and hydropic degeneration. Focal areas of hemorrhages and necrosis were also seen in liver and kidneys. Vascular lesions in the form of congestion, thrombosis and necrosis of blood vessels as well as perivascular mononuclear infiltration were also pronounced. The mostly affected organs were brain and testes especially in rats given dimethoate contaminated feed. These changes included vesiculation in the brain tissue, encephalomalacia as well as satellitosis and neurophagia. The testes showed atrophy of seminiferous tubules together with fibrosis, intertubular edema and failure of spermatogenesis. We advise that great attention should be taken when dealing with such insecticides in order to avoid its various adverse action on different body tissue.
of farm animals or human in contact.

Introduction

The wide application of organophosphorus insecticide in agriculture represents a great hazard to livestock. As the prolonged exposure to these contaminants even at low concentrations may lead to toxicity, immuno-suppression or reproductive failure (Nafstad et al., 1983). In Egypt, Dogheim et al., (1996) recorded organochlorine and organo-phosphorus pesticides, including those have been prohibited from use, in human milk and environmental samples collected from Kafr El-Zayat governorate. Dimethoate is an organophosphorus insecticide used extensively in agriculture as a systemic insecticide and acaricide for gardens, vineyards and field crops (Humphreys, 1988). The toxic effects of dimethoate were studied by many authors. Metelov et al., (1977) stated that dimethoate have a neurotoxic effect in sheep, calves and fish. In human being, Krieger and Thongsinthusak, (1993) found that dimethoate is readily absorbed and its urinary metabolites are readily eliminated following to low doses. Affi et al., (1991) recorded a dose-related decrease in the weight of the most genital organs in male albino rats. Also reduced sperm motility associated with increase in the percentage of dead and abnormal spermatozoa of treated rats. Level of plasma testosterone was lowered in treated groups and histological examination revealed that dimethoate caused moderate to severe degenerative changes of spermatogonial cells. The highest tissue residues of dimethoate were recorded in liver and testes and the lowest was recorded in skeletal muscles. Institots et al., (1995) studied the immunotoxicity of repeated doses of dimethoate and methyl parathion given to rat over three generations and found that dimethoate had a detectable effect on body weight, birth weight and number, organ weight, hematological parameters and immune function. Sirvastava and Raizada, (1996) recorded that dimethoate produced enzymatic changes in liver of rat associated with pathomorphological changes in liver and brain. They also noticed reduced acetyl choline esterase activity in fetal brain and placenta when given to dams indicating possible transmigration of dimethoate from dams to fetuses.

Concerning to persistency of dimethoate, Pareek and Kavadia, (1988) indicated that the waiting period for dimethoate was (5 to 6) days to avoid consumer risk after its application at 0.03% on musk melon, long melon, and ridge gourd. On the same aspect, Cabras et al., (1995) studied the persistency of some organophosphorus compounds on orange fruits and found that the residues of dimethoate was only found in the fruit peel and a very low concentrations were detected in the fruit pulp.

The current study was undertaken to study the toxopathological effect of dimethoate at different concentrations and to evaluate its effect on fertility of male albino rats. Furthermore to detect the tissue residues of dimethoate after direct admini-
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istration to rats or when sprayed on edible plants and given to rats.

MATERIAL AND METHODS

Materials:
Chemicals: Dimethoate * El-Naser. 2 * an organophosphorus Insecticide was obtained as emulsifiable liquid concentration containing 40 % ( W/V ) active ingredient ( product of El-Nasar Co. Egypt ). Its chemical names is O.O dimethyl S-(N- methyl) carbamoyl methyl phosphorodithioate.
LD$_{50}$ of dimethoate in rats is 250 mg / kg B. wt. ( Ware , 1978).

Animals , dosing and grouping: One hundred and twenty mature albino rats of both sexes weighing 150-160 gm B. wt. were divided into two main groups

Group A:
One hundred rats were divided into four subgroups a, b, c, d, each of 25 rats of both sexes. The animals were given daily doses of dimethoate diluted in distilled water at concentrations of 1/ 40 1/ 20 and 1/10 of LD$_{50}$ by stomach tube to the 1st three treated groups respectively. The fourth group was given distilled water and kept as control. Five rats from each subgroup were sacrificed after one, two, three and four weeks for histopathology and serum was collected at the end of the experiment " 4 weeks" for some biochemical analysis and blood was collected on anticoagulant for determination of choline esterase activity.

Group B:
Twenty male rats were classified into 4 subgroups each of five rats, fed on a green forage sprayed with dimethoate at the concentration indicated in the pamphlet and offered to animals after 2, 4, 8 days of application in the 1st three treated subgroups respectively. The other control subgroup offered dimethoate free green forage.

Methods:
For animals in group A:- serum samples were separated for determination of some biochemical parameters as follows:

* Serum total protein , albumin and globulin after Welchelbaum ( 1945), Daumais et al., (1971) and Coles , (1986) respectively.

* Alkaline phosphatase was estimated using method of Kind and King ( 1954).

* Serum transaminases ( GPT and GOT ) were estimated by the method of Reitman and Frankel ( 1957) and serum urea and creatinine were determined according to Hundan and Rapoport , (1968) and Chaney and Marbach , (1963) respectively.

* Cholinesterase was determined in whole blood using reagent kit of BioMarieux , according to Whittaker , (1984).

Method adopted in group B:
Semen evaluation:
This group offered green forage sprayed with dimethoate and offered to animals after 2, 4 and 6 days of application. Feeding continued for 65 days to cover the period of spermatogenic cycle which ranges from 56 - 60 days in rats ( Hershberger et al., 1999) then animals were scarified and the testes , seminal vesicles and prostate

glands were dessicated and weighed. The cauda epididymides were minced in normal saline and a drop of this epididymal suspension was picked up for semen evaluation according to Zemjanis, (1970).

Testosterone estimations:
At the end of the experiment, serum was collected for estimation of testosterone by enzyme immunoassay and reagent kits supplied from Orion Diagnostica, Finland. Samples were measured for testosterone using ELISA reader.

Tissue residues:
After slaughtering, samples from testes, liver, and skeletal muscles were collected from each animal. One gram of each tissue was thoroughly homogenized in absolute alcohol at a rate of 1 g of tissue to 10 ml alcohol, then centrifuged at 3,000 r.p.m. for 15 minutes. The supernatant was filtered through 0.45 µm filter paper and transferred to clean tubes and used for estimation of dimethoate concentration using high performance liquid chromatography with programmable u.v detector.

It was chromatographed on a reverse-phase C18 column. The appropriate volumes of samples were injected into column and retention time were measured at 254 nm with the mobile phase of 50:20 acetonitrile and water. (Zebra et al., 1995).

Histopathology:
Specimens from liver, brain, heart, kidney, lung and testes were taken and fixed in 10% buffered neutral formalin solution. After proper fixation, these specimens were processed and then sections 5 microns thickness and stained using hematoxylin and eosin after (Drury and Wallington, 1980).

Statistical analysis was carried out according to Sendecore, (1971).

RESULTS

Table (1) revealed that choline esterase activity decreased due to dimethoate at a dose related pattern. A similar pattern have been recorded for the percentages of total protein, albumin, globulin while GPT, GOT, alkaline phosphatase, urea and creatinine showed a dose dependent increase. Concerning to the effect of dimethoate given with green forage to rats after 2, 4, 6 days of spraying and continued for 65 consecutive days table (2) revealed reduction in the weight and testicle, seminal vesicle and prostate gland and this decrease was correlated with the time elapsed from spraying.

Table (3) indicates a decreases in sperm concentration, percentages of live sperm and motility due to green forage contaminated with dimethoate. Also the level of testosterone nmol/L decreased while the percentages of total sperm abnormalities increased compared with the control group.

Table (4) indicate tissue residues of dimethoate in liver, testes and skeletal muscles and higher in liver followed by testes and the lowest residues recorded in skeletal muscles. Also the table indicates a correlation between tissue residues and time elapsed after spraying.

Concerning to clinical signs, group A, showed
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Table 1: Effect of dimethoate on some biochemical parameters at different dose levels (1/40 LD_{50}, 1/20 LD_{50}, 1/10 LD_{50}) in serum of albino rat after administration for 1 month.

<table>
<thead>
<tr>
<th>Biochemical parameters</th>
<th>Control</th>
<th>1/40 LD_{50}</th>
<th>1/20 LD_{50}</th>
<th>1/10 LD_{50}</th>
</tr>
</thead>
<tbody>
<tr>
<td>Choline esterase activity (U/L)</td>
<td>3262.25±22.91</td>
<td>2153.5±24.90</td>
<td>1925.62±22.81</td>
<td>1668.12±21.63</td>
</tr>
<tr>
<td>Total protein (gm %)</td>
<td>8.1±0.22</td>
<td>7.20±0.20</td>
<td>6.50±0.21</td>
<td>5.60±0.20</td>
</tr>
<tr>
<td>Albumin (gm %)</td>
<td>4.20±0.25</td>
<td>4.00±0.26</td>
<td>3.90±0.23</td>
<td>3.80±0.25</td>
</tr>
<tr>
<td>Globulin (gm %)</td>
<td>3.90±0.23</td>
<td>3.20±0.21</td>
<td>2.60±0.22</td>
<td>1.80±0.20</td>
</tr>
<tr>
<td>GPT (U/ml)</td>
<td>72.41±1.53</td>
<td>85.22±1.75</td>
<td>95.20±2.35</td>
<td>102.40±2.55</td>
</tr>
<tr>
<td>GOT (U/ml)</td>
<td>84.44±2.29</td>
<td>92.65±2.10</td>
<td>102.22±1.80</td>
<td>112.02±1.85</td>
</tr>
<tr>
<td>Alk. phosph. (U/100 ml)</td>
<td>24.20±0.60</td>
<td>32.22±0.66</td>
<td>33.32±0.80</td>
<td>35.21±1.20</td>
</tr>
<tr>
<td>Urea (gm %)</td>
<td>20.23±2.51</td>
<td>33.21±2.66</td>
<td>55.72±2.71</td>
<td>120.14±2.62</td>
</tr>
<tr>
<td>Creatinine (gm %)</td>
<td>2.12±0.62</td>
<td>2.40±0.91</td>
<td>4.12±0.92</td>
<td>5.20±0.90</td>
</tr>
</tbody>
</table>

Table 2: Effect of dimethoate on relative body weight (gm / 100 gm B. wt) of male reproductive organs after administration with green forage for 65 days after 2, 4 and 6 days of spraying (mean ±S.E).

<table>
<thead>
<tr>
<th>organ</th>
<th>control</th>
<th>6 days after spraying</th>
<th>4 days after spraying</th>
<th>2 days after spraying</th>
</tr>
</thead>
<tbody>
<tr>
<td>Testicle</td>
<td>1.5 ± 0.012</td>
<td>1.33 ± 0.029</td>
<td>1.07 ± 0.018</td>
<td>0.97 ± 0.016</td>
</tr>
<tr>
<td>Seminal vesicle</td>
<td>0.22 ± 0.008</td>
<td>1.33 ± 0.029</td>
<td>0.12 ± 0.006</td>
<td>0.10 ± 0.007</td>
</tr>
<tr>
<td>Prostate gland</td>
<td>0.18 ± 0.007</td>
<td>0.14 ± 0.006</td>
<td>0.08 ± 0.006</td>
<td>0.07 ± 0.006</td>
</tr>
</tbody>
</table>
Table (3) Effect of dimethoate on semen picture and serum testosterone of male albino rat after administration with green forage at different period of application for 65 consecutive days.

<table>
<thead>
<tr>
<th>Semen picture</th>
<th>control</th>
<th>6 days after spraying</th>
<th>4 days after spraying</th>
<th>2 days after spraying</th>
</tr>
</thead>
<tbody>
<tr>
<td>Progressive motility %</td>
<td>82 ± 1.31</td>
<td>66 ± 2.1***</td>
<td>51 ± 2.3***</td>
<td>46 ± 2.6***</td>
</tr>
<tr>
<td>Live sperm %</td>
<td>85 ± 0.95</td>
<td>65 ± 3.2***</td>
<td>52 ± 2.2**</td>
<td>49 ± 2.1**</td>
</tr>
<tr>
<td>Sperm Con. (X 106/ml)</td>
<td>316.61 ± 10.97</td>
<td>296.14 ± 10.12**</td>
<td>275.42 ± 8.75**</td>
<td>260.14 ± 7.61**</td>
</tr>
<tr>
<td>Total sperm abnormalities (%)</td>
<td>1.50 ± 0.30</td>
<td>8.2 ± 0.62**</td>
<td>16 ± 0.78**</td>
<td>20 ± 0.78**</td>
</tr>
<tr>
<td>Serum testosterone nmol/L</td>
<td>3.80 ± 0.32</td>
<td>3.25 ± 0.41**</td>
<td>2.52 ± 0.35</td>
<td>2.31 ± 0.55</td>
</tr>
</tbody>
</table>

Table (4) Dimethoate residue (PPM) in tissue of albino rat after its administration with green forage and given to animal after (2, 4, 6) days of application for 65 consecutive days. (mean ± S.E).
N.D = not detected

<table>
<thead>
<tr>
<th>Sample</th>
<th>control</th>
<th>6 days after spraying</th>
<th>4 days after spraying</th>
<th>2 days after spraying</th>
</tr>
</thead>
<tbody>
<tr>
<td>Liver</td>
<td>N.D</td>
<td>1.52 ± 0.07</td>
<td>2.44 ± 0.07</td>
<td>2.91 ± 0.08</td>
</tr>
<tr>
<td>Testes</td>
<td>N.D</td>
<td>0.87 ± 0.04</td>
<td>1.25 ± 0.06</td>
<td>1.62 ± 0.06</td>
</tr>
<tr>
<td>Muscle</td>
<td>N.D</td>
<td>0.32 ± 0.02</td>
<td>0.61 ± 0.03</td>
<td>0.92 ± 0.03</td>
</tr>
</tbody>
</table>

N.D = not detected
livation, lacrimation, tremors, watery diarrhea and partial loss of appetite after one and two weeks from administration in all sub-groups. Meanwhile, after three and four weeks, severe emaciation, roughened hair, excitation and loss of appetite were prevalent. The severity of signs tolerated by increasing the dose and time of exposure.

In group B, gradual emaciation, loss of hair and incoordination were the most clinical signs recorded.

**Pathological findings:** Post-mortem examination of rats received 1/10 LD50 revealed severe congestion of liver, kidneys, heart and brain with the presence of petechiae on their surfaces after one and two weeks from administration. Meanwhile, after three and four weeks, diffuse areas of hemorrhages with the presence of grayish white foci on the surface of liver, heart and kidneys were seen. The testes were congested and small in size (Fig.1).

Microscopically, after one and two weeks from administration, the liver showed severe congestion of blood vessels and sinusoids. Focal areas of hemorrhages with perivascular mononuclear cellular infiltration. The hepatocytes were suffered from mild degenerative changes in the form of vacuolar and hydropic degeneration (Fig.2). The kidneys showed congestion of the renal blood vessels and intertubular blood capillaries. The glomeruli showing congestion of the glomerular tuft. Periglomerular hemorrhages were also seen (Fig.3). The heart and brain showed congested blood vessels and focal areas of hemorrhages. Meanwhile, after three and four weeks, focal areas of necrosis in the form of structureless eosinophilic substances infiltrated with mononuclear cells were detected in the liver and kidneys. The heart showed focal mononuclear cellular aggregations. The testes showed degenerative changes in the primary and secondary spermatocytes with the presence of sperm giant cells in the tubular lumen (Fig. 4). The rats received a dose of 1/20 of dimethoate LD50 showed congestion of parenchymatous organs with the presence of petechiae on their surfaces after one and two weeks from administration. But after three and four weeks the brain also showed severe congestion of blood vessels and the testes were congested and somewhat decreased in size. Microscopically, after one and two weeks severe congestion, hemorrhages and vascular thrombosis were seen in the liver, kidneys and heart (Fig.5). Moreover, the kidneys showed intertubular mononuclear cellular infiltration (Fig.6). The renal tubules showed degenerative changes in the form of cloudy swelling and vascular degeneration. The heart showed focal areas of myelomalacia infiltrated with mononuclear cells (Fig.7). The testes showed intertubular edema with congestion of intertubular blood vessels (Fig.8). The brain showed vesculation in the brain substances with focal gliosis after three and four weeks (Fig. 9 & 10). The rats received 1/40 LD50 showed congestion of the liver, kidneys and heart with the presence of petechial hemorrhages on their surfaces. The brain showed congestion of its blood vessels. These gross findings
increased by atime of experiments. Microscopically, congestion, hemorrhages with perivascular mononuclear infiltration in the liver and kidneys were prevalent. Moreover, heart showed intermuscular hemorrhage. By atime the renal tubules showed degenerative changes in their epithelial cell lining with the presence of casts in the tubular lumina (Fig.11). The brain showed perivascular mononuclear infiltration. The testes showed congestion of testicular blood vessels.

**Group B:** The post mortem examination showed a marked decrease in the parenchymatous organs. Grayish white foal were detected in the liver, kidneys and heart. Some examined cases showed paleness or yellowish coloration of the liver and kidneys. The brain showed petechial hemorrhages. The testes were markedly decreased in size and become firm with thickened and wrinkled capsule. Microscopically, the liver showed congestion of blood vessels and sinusoids with focal mononuclear Infiltration.(Fig.,12). The portal areas showed fibrous tissue proliferation together with hyperplasia of the epithelial cell lining the bile duct with newly formed bile duct. Portal mononuclear infiltration were also detected. The kidneys showed presence of hyaline casts in the renal tubules with focal areas of necrosis infiltrated with mononuclear cells. Focal replacement of the renal tissue by fibrous connective tissue (Fig,12). The heart showed focal areas of myelomalacia and hemorrhages. The brain showed multiple areas of encaphalomalacia with satellitosis and neuropathia of the neuron.(Fig., 13). Diffuse hemorrhage in brain was also detected (Fig,14). The testes showed atrophy of the seminiferous tubules with complete absence of spermatogonia (Fig., 15). Intertubular edema and fibrous tissue proliferation were also detected (Fig., 16).

**DISCUSSION**

Pesticides are double edged weapon, they are widely used to control pests which led to 30 - 40% losses in crops all over the world. However, they leave some residues in crops and are incriminated in environmental pollution. Many investigations have been dealt with the harmful influence of insecticides used for controlling of animal parasites, but few studies were done on plant insecticides to find out their hazards in animals which may consume forage recently treated by such pesticides. The toxic effect of these pesticides may be greatly reflected on productivity and reproductivity of farm animals beside its immune potents against prevalent diseases. The current study deals with an organophosphorus compound (dimethoate) with the aim of exploring its effect on reproduction, pathological effects and its residues.

Table (1) shows the effect of dimethoate on some biochemical parameters of serum in intoxicated animals. Like other organophosphorus compound it was expected that cholinesterase activity will reduced due to dimethoate. Similar results were recorded by Metelov et al., (1977) due to dimethoate in fish; Hamza et al., (1991) due to carbofuran, carbamate insecticide; and by El-
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Chromatogram (1): Showing residues of dimethoate in liver (A), testes (B), and muscle (C) after 65 days of feeding contaminated forage offered to animals after two days of application.
Chromatogram (2): Showing residues of dimethoate in liver (A), testes (B), and muscle (C) after 65 days of feeding contaminated forage offered to animals after four days of application.
3: Showing residues of dimethoate in liver (A), testes (B), and muscle (C) after 65 days of feeding contaminated forage offered to animals after six days of application.
t al., (1993) due to confider organophosphorus compound in albino rat and it was also recorded in fetal brain and placenta when dimethoate was given to dams in albino rat (Srivastava and Raizada, 1996). A pronounced reduction was recorded due to dimethoate on the levels of total protein and globulin with no remarked effect on the level of albumin. The decrease of total protein may be due to damage effect on protein biosynthesis machinery and or inhibition to m RNA transcription (El-Sheikh et al., 1993).

The effect of dimethoate on the enzyme indices of liver function shows a remarkable dose related increase in the activity of GOT, GPT and alkaline phosphatase. It is known that transaminases are important enzymes in all biological processes as they found mainly in liver and their level in blood serum is a good indicator for diseased and damaged liver (Wilkinson, 1970 and Garb, 1971).

Our results concerning the pathology confirmed this damage in liver (Fig. 2, 5) which agree the results recorded by Hamza et al., (1991) due to carbofuran, carbamate insecticide. However and in contrast to the present investigations, El-Sheikh et al., (1993) recorded that a single dose of confider caused a gradual decrease in both GOT and GPT and also disagree with the result of Elia and Bayomy (1992) using cyanophos pesticide in albino mice. This alteration may be due to difference in potency and structure of examined pesticide and may also be due to various experimental condition. As indices for kidney function, urea and creatinine levels showed a persistence elevation with the dose given. This finding indicates damaging effect of dimethoate to the kidney a process that confirmed in our histopathological investigations (Figs. 3, 6, 12).

The effect of dimethoate on the relative weight of male reproductive organs after administration with green forage for 65 consecutive days and at various period of application on plant is shown in table (4). Plant sprayed with dimethoate reduced the weight of testicle, seminal vesicle and prostate gland at various period of application (2, 4, 6 days). The reduction was more pronounced at fewer period after application on plant. This picture indicated that the insecticide is metabolised by time, any how it was still effective during the examined period after application. The recorded decrease in the weight of sexual organs may be attributed to a direct destructive effect of dimethoate on male sexual organs especially tests as recorded in our study (Figs. 4, 8, 15, 18). In this respect our results agree that recorded previously by Afifi et al., (1991) and Institoris et al., (1995).

Table (3) shows another indication for the role of dimethoate on male fertility where the deleterious effect of dimethoate given with green forage was pronounced and manifested by reduction in the percentages of progressive motility, live sperm and sperm concentrations synchronized with detectable increase in the percentage of total sperm abnormalities. The alterations in sperm picture in treated animals were inversely correlated with the time elapsed of dimethoate spraying on the
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green forage.
The reduction in sperm cell concentration could be attributed to reduction in meiotic index of testicular cells as being recorded for pyrethroid in albino rat (El-Ashmawy et al., 1993). The manifestation of testis in our histopathology may add another explanation or the present status of semen picture. Our results shows a complete agreement with pervious studies dealt with pesticide e.g. Affi et al., (1991); Hamza et al., (1991) and El-Ashmawy et al., (1993) for dimethoate, carbofuran and Matox (pyrethroid), respectively. Table (5) shows also that serum testosterone reduced in the groups fed on green forage sprayed with dimethoate. The decrease in the level of testosterone might explain the significant depression in the testicular, epididemis, seminal vesicle and prostate gland relative weight and a significant reduction in sperm concentrations previously explained by El-Ashmawy et al., (1993). Our results coincided with that recorded by Affi et al., (1991).

Table (4) and chromatograms (18.2&3) shows the residues of dimethoate in tissue of albino rats given in green forage at different period of spraying for 65 consecutive days. Liver showed the highest concentrations of dimethoate followed by testes and the lowest concentrations was recorded in skeletal muscles. Our data means that dimethoate is stable compounds and leave various concentrations in (liver, testes and skeletal muscles) even after 8 days of application on plants. In this respect our results agree with metlov et al., (1977) who found dimethoate as a parent compound in fish tissues for 40 days and attributed the remained concentration in fish to the reduction in acetyl choline esterase activity in the brain. Also we refer that Affi et al., (1991) recorded different residues due to dimethoate in albino rat. However, Krieger and Thorgsthusak, (1993) stated that dimethoate is readily absorbed and its urinary metabolites are readily eliminated following to low doses. We have to mention also that Pareek and Kavadia, (1988) indicated that the waiting period for dimethoate was 5-6 days to avoid consumer risk. This work seems to be in agreement with our findings.

Concerning to histopathological investigations our results revealed that the administration of different concentrations of dimethoate had adverse effect on various body tissues according the dose and time of administration. As we found that most of parenchymatous organs (liver, kidneys and heart) showed degenerative changes, hemorrhage and focal areas of necrosis. These findings were in complete agreement with Abol-Gharr, et. al., (1963), Krasuse and Homola (1974); Clark and Clark, (1975); El-Mansoury, (1983); ElSwak, (1989); El-Sawak, (1990); Affi et al., (1991) and El-Swak, et al., (1992) who found similar changes in some organophosphorus insecticides. Meanwhile, testicular changes in our results were considered the most important lesions especially in rats given the green forage sprayed with dimethoate at different period of application and given for 65 successive days. The testes were small in size, tense and showed wrinkled capsule. Microscopically, degenerative

*Alex. J. Vet. Sci., Vol. 13, No. 2, October 1997*
changes in primary and secondary spermatocytes with complete absences of spermatogenesis were seen. Moreover, atrophy of seminiferous tubules with fibrous tissue proliferation were detected. These findings were in a partial agreement with that mentioned by Jackson and Jones (1968); Krause and Homola, (1977) and Affifi et al., (1991) who found degenerative changes and hemorrhage in the testes of rats received dimethoate. These degenerative changes especially in the reproductive organs returned to the infertility action of dimethyl groups as previously mentioned by Affifi et al., (1991). Our results revealed that the nervous manifestation in rats received dimethoate at different concentrations as confirmed microscopically by congestion of brain blood vessels, perivascular mononuclear infiltration together with multiple areas of encephalomalacia, satellitosis and neurophagia. These findings were in a partial agreement with that mentioned by Abbasy et al., (1989) who found fragmentation of axons, degeneration in neurons as well as demyelination of sciatic nerve in cases of sulprofos treated hens.

In conclusion, the current study proved that dimethoate is an organo-phosphorous compound has toxopathological action especially on brain and testes leading to male reproductive failure and leave residues in animal tissue with predicted consumer risk. Therefore great attention should be taken during field application of dimethoate and similar insecticides to avoid the possible adverse action on various body tissue of farm animals and occupationally exposed human.

List of figures:

(Fig. 1) Showing a marked decrease in size of testes of rats received 1/10, 1/20 and 1/40 from LD₅₀ after four weeks from administration. A control, B, 1/10 C, 1/20 D 1/40 LD₅₀.

(Fig. 2) Liver of rat received 1/10 LD₅₀ after two weeks showing congestion of central vein and vacuolar degeneration of hepatocytes. (H & E stain X 600)

(Fig. 3) Kidney of rat received 1/10 LD₅₀ after two weeks showing intertubular hemorrhage. H & E stain X 250

(Fig. 4) Testes of rat received 1/10 LD₅₀ after four weeks from administration showing degenerative changes in spermatocytes with presence of sperm giant cells in their lumen. (H & E stain X 450)

(Fig. 5) Liver of rat received 1/20 LD₅₀ after two weeks from administration showing intermuscular hemorrhage and hyalinization of the cardiac muscles. (H & E stain X 630)

(Fig. 6) Kidney of rat received 1/20 LD₅₀ after two weeks showing degenerative changes in the renal tubules together with Intertubular
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mononuclear infiltration. (H & E stain X 600)

(Fig. 7) showing focal area of myomalacia with mononuclear infiltration in the heart of rat received 1/20 LD₅₀ of dimethoate after four weeks. (H & E stain X 400)

(Fig. 8) Testes of rat received 1/20 LD₅₀ after four weeks showing intertubular edema and degeneration of seminiferous tubules. (H & E stain X 630)

(Fig. 9) showing vesiculation of the brain substance in rat received 1/20 LD₅₀ of dimethoate after four weeks from administration. (H & E stain X 450)

(Fig. 10) Brain of rat received 1/20 LD₅₀ after four weeks showing focal area of gliosis. (H & E stain X 450)

(Fig. 11) Kidney of rat received 1/40 LD₅₀ of dimethoate after four weeks from administration showing degeneration and cast formation in the renal tubules.
(H & E stain X 300).

(Fig. 12) Kidney of rat fed on dimethoate sprayed forage for 65 days by showing focal fibrosis of the renal tissues. (H & E stain X 450)

(Fig. 13) Brain showing encephalomalacia in rat received forage sprayed by dimethoate and fed for 65 days. (H & E stain X 300)

(Fig. 14) Brain showing focal area of hemorrhage in rat fed on dimethoate sprayed forage by for 65 days. (H & E stain X 350)

(Fig. 15) Testes of rats showing atrophy of seminiferous tubules which fed on dimethoate sprayed forage for 65 days. (H & E stain X 300)

(Fig. 16) showing degeneration in the epithelial lining the seminiferous tubules in rat fed on dimethoate sprayed forage for 65 days. (H & E stain X 450)

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التأثير الباثولوجي للتسمم بالدييثيوت وتأثيره على الظهورة ومتبقياته في أنسجة الفقار البيضاء

د. هشام محمد أبو صافر

استهدف هذا الدراسة تأثير البيع العشري فى السنو (دييثيوت) على النكتة التامانية، classe et متبقياته في الأنسجة وكذلك تأثيره الباثولوجي حيث تم إعطاء الميدين بجرعات 1200 و 600 مللي جرعة نصف المبتيلة مباشرة عن طريق الفم كما تم رش الميا على علائق حيوية واعتدالات النمو دورة 15 يومًا متتالية.

وقد أظهرت النتائج أن الديثيوت أدى إلى تغيير المناعات الكيميائية لمصل الفقار مما يؤدي إلى تأثير وظائف الكبد و الكلى. هذا بالإضافة إلى أن صورة السائل المذوء و وزن الانضغاط الدموية (الحبيبة والروستم) والتحريرات الملونة) مع انخفاض معدل النسيج وتراجع موثر على انخفاض التكلفة الناقصة. كما ظهرت متبقيات للميدين في النكتة الكبد والكبد والبيض.

كما أظهر النقص الباثولوجي ووجود ثروات سادية في الكبد والكلى والقلب على طية جزء فيباد واستحالات مانية. كما ظهرت مواصلات رفيعة وتكون في الكبد والكلى، ووحشي اضطرابات وانضغاطات وتطار في الأوعية الدموية مع انتشار بعض الخلايا الاستجابة. وقد أفادت النتائج إلى أن الميدين التجانيين كانت أكثر الأعضا تأثيرا خاصة في الفقار الذي أظهرت غذا ملثما بالبيده حيث ظهرت متابعة تندر من الاعتدالات النموية في الخلايا المذعة بجهة تنوعات و تأثيات و انسجام في إنتاج الحيوانات المنوية.

وقد يتبين بصحة العمرية القائمة عند تداول هذا البيدي لتجنب آثاره المختلفة على النكتة.