

The Results

Table (1) and fig. (1):-

Show mean \pm standard deviation of age (in years) of the three groups of our study. Statistical analysis showed no significant difference between septic group and control group.

Table (2) and fig. (2):-

Show percentage (%) of sex distribution of the three groups. There was no statistical difference between the three groups.

Table (3) and fig. (3):-

Show percentage (%) of clinical manifestations in-patients of septic and aseptic groups. Neck stiffness was the most common presentation and coma was the least presentation

Table (4):

Shows C.S.F. picture of the three groups. All cases of septic meningitis show turbid aspect of C.S.F while C.S.F of a septic and control groups are clear. There was marked hypercellularity of C.F.F of septic and aseptic group with predominance of polymorphnuclear pleocytes in septic group and mononuclear pleocytes in aseptic group.

In contrast with aseptic and control groups, C.S.F of septic group shown significant increase in protein (mean = $215.79 \pm \text{SD} = 83.89$ mg%) and significant decrease in glucose (mean = $14.71 \pm \text{SD} = 4.94$ mg%).

Table (5):-

Shows blood picture of the three groups there is significant decrease of HB % (mean = $9.72 \text{ gm/ } 100\text{ml}$) and significant leucocytosis (mean = $9.60 \text{ thousands/mm}^3$) in septic group as compared to aseptic and control groups).

Table (6): and fig. (4): -

Shows the levels of NO in C.S.F of the three groups before starting treatment, there is significant elivation of NO level in C.S.F of septic group (Mean = $23.13 \pm \text{SD} = 6.58 \text{ nmol/ml}$) as compared to its levels in aseptic (Mean = $7.55 \pm \text{SD} = 2.99 \text{ nmol/ml}$) and control groups (Mean = $8.02 \pm \text{SD} = 3.76 \text{ nmol/ml}$).

Table (7) and fig. (5):-

Shows the level of NO in serum of the three groups before starting treatement, there was significant elivation of NO level in septic group (mean = $83.23 \pm \text{SD} = 17.60 \text{ nmol/ml}$) as compared to its levels in aseptic (mean = $66.41 \pm \text{SD} = 26.90 \text{ nmol/ml}$) and control groups (mean = $42.31 \pm \text{SD} = 6.22 \text{ nmol/ml}$).

Table (8):-

Shows correlation between NO and other variables in septic group, there was significant positive correlation between cells and protein in C.S.F and NO level in serum and C.S.F. Also, there was significant negative correlation between glucose and chloride in C.S.F and NO level in serum and C.S.F. However, there was no significant correlation between NO in serum and C.S.F. and other variables as fever, Hb, RBCs, WBCs, platelets.

Table (9):-

Shows correlation between age (in years) and NO (in nmol/ml) in control group, there was no significant correlation between age and NO.

Fig. (6):-

Shows the distribution of organisms in septic groups as shown by the result of culture and sensitivity of C.S.F of septic groups, the commonest organism was *H. influenzae* (65%), the other organisms was *N. meningitidis* (21%) and *St. pneumonia* (14%).

Table (10) and fig. (7):-

Shows the relation between caustive organisms and NO level (in nmol/ml) in serum and C.S.F of septic group. It was found that NO is more elivated in cases with *H.influenzae* than cases with *St.pneumonia* or *N.meningetidis*.

Table (11) and fig. (8):

Shows comparison between NO (in nmol/ml) pretreatment and posttreatment in septic group in both serum and C.S.F. There is significant decrease of NO level in both serum and C.S.F after treatment with antibiotics and dexamethasone.

Fig. (9):-

Shows prognosis in studied groups, About 20% of cases of septic group with bad prognosis and about 80% with good prognosis. 100% of cases of aseptic group with good prognosis.

Table (12) and fig. (10):-

Shows the relation between prognosis and NO (in nmol/ml) in septic group, there is significant increase in NO levels in cases with bad prognosis than cases with good prognosis.

microvascular damage, brain edema formation and C.S.F. pleocytosis (*Boje, 1995, and Buster et al., 1995*).

NO is a short-lived free radical produced by a variety of cell types and involved in physiologic processes, such as smooth muscle relaxation, neuronal signaling, inhibition of platelet aggregation and regulation of cell mediated cytotoxicity (*Anggard, 1994, Bredt et al., 1994, Lowenstein et al., 1994 and Moncada et al., 1993*).

The cellular site of NO production in children with bacterial meningitis is unknown. However, NO can be generated by a variety of cell types, including neutrophils, microglia/macrophages, endothelial cells, astrocytes, neurons and vascular smooth muscle cells and by bacteria.

In the brain, NO has several important function and effects including neurotransmission, regulation of cerebral vascular tone and cerebral blood flow, and mediation of ischemic and excitotoxic neuronal injury (*Wang and Ferriereo et al., 1995*).

Intracisternal inoculation of heat-killed bacteria induces an increase in C.S.F. nitrite comparable to that observed with living bacteria (*Koedel et al., 1995*).

Buster et al., 1995 demonstrated that inoculation of live bacteria (*Hemophilus influenzae* type b) or lipopolysaccharides intracisternally in rats resulted in a rise of C.S.F NO and increased the permeability of blood-C.S.F barrier. These alterations were inhibited by systemic administration of N-nitro-L-arginine methyl ester, a non-selective NO synthase inhibitor.

Bernatowicz et al., 1995 demonstrated that Co-incubation of rat astrocytes, microglial cells and neurons in primary cultures with heat killed uncapsulated streptococcus pneumoniae stimulates NO production, a process inhibited by NO synthase inhibitors (including aminoguanidine), cycloheximide and dexamethazone.

Our study was carried out on (40) childrens from Benha fever hospital, they were divided into three groups:

Group (A): included 14 childrens with septic meningitis. Their ages ranged from 6 months to 9 years. They were proved to be septic meningitis by clinical picture of meningitis (e.g. fever, convulsions, vomiting, bulging anterior fontanelle, necks and back stiffness, kerning's sign, Brudzinski's signs, drowsiness or coma). (Table 3 and fig. 3). The presence of highly elevated C.R.P, polymorphnuclear pleocytosis in C.S.F and positive C.S.F bacterial cultures where *H.influenzae* was commonest organism (65%), the other organisms were *Niesseria meningetdis* (21%) and *St. pneumonia* (14% of cases).

Group (B) Included 16 children with aseptic meningitis their ages ranged from 6 months to 8 years. They were proved to be aseptic meningitis by clinical picture of meningitis. (**Table 3 and fig. 3**), negative or slightly elevated C.R.P., pleocytosis in C.S.F. and the presence of negative bacterial cultures of C.S.F.

Group (C) Included 10 children as a control group, their ages ranged from 6 months to 10 years they were admitted to Benha fever hospital presented with fever and required C.S.F analysis to exclude meningitis, they were later found to be free of meningitis as evidenced by the results of C.S.F examination and negative bacteriological culture of C.S.F. and negative clinical findings of meningitis during follow up.

All studied groups and neurological examination, C.B.C, C.R.P and C.S.F. analysis (physical, cellular, chemical, culture and sensitivity), the results of C.S.F analysis revealed significant elevation of cells in cases of meningitis with predominance of polymorphnuclear pleocytes in septic meningis and predominance of mononuclear pleocyte in cases of aseptic meningitis, also there was significant increased protein and decreased C.S.F glucose in cases of septic meningitis. As regard aspect there was turbidity of C.S.F in cases of septic meningitis as compared to clear C.S.F in cases of aseptic meningitis and control group.

In the present study we measured the level of NO in serum and C.S.F in the three groups before treatment, and also its level after

treatment in septic group. There was a significant increase in level of NO in both serum and C.S.F in children with septic meningitis as compared with those of a septic and control groups. These results provide evidence of an enhanced production of NO in C.S.F and serum of children with septic meningitis.

Our results are in agreement with those of *Pfister et al., (1995)* who reported that there was increased C.S.F levels of NO metabolites in patients with bacterial meningitis. Also, in agreement with the results of *Furth et al., (1995)* and *Kornelisse et al., (1996)* who documented a significant elevation of C.S.F NO in children with bacterial meningitis caused by *H.influenzae*, *N.meningitidis* or *St. pneumoniae*.

Our results did not show any significant correlation between age and NO level in serum and C.S.F among the control group as shown in table (9) also, we found a significant correlation between serum and C.S.F levels of NO and glucose and protein levels in C.S.F among patients with bacterial meningitis as shown in table (8).

NO may inhibit the mitochondrial respiration that enhances anaerobic glycolysis. This mechanism may contribute to the decrease glucose concentration in C.S.F compartment in patients with bacterial meningitis (*Albina JE, et al., 1993* and *Welsh N, et al., 1992*).

Alternatively, low glucose level may also be explained by inhibition of carrier-mediated transport across B.B.B. (*Cooper A, et al., 1968* and *Prockop L, et al., 1966*).

As regard organism in C.S.F of septic group in our study the result of the culture of C.S.F revealed three types of bacteria; H.influenzae (65%), N.meningitidis (21%) and St. pneumonia (14%) (**fig. 6**).

There was a significant increase in C.S.F and serum NO in H.influenzae meningitis as compared with those of meningitis caused by St.pneumoniae and by N.meningitidis thus, H.influenzae may be a more potent inducer of inducible NO synthase than N.meningitidis and St.pneumonia, these results agreed with those of *Tsukahara et al., (1988)*.

In our study, two patients with meningitis caused by H.influenzae died during treatment and follow up and a third child with H.influenzae meningitis developed neurological complication. It was noted that NO was more elevated in C.S.F and serum in those patients than other children with meningitis. This suggests that NO has a role in the prognosis of these cases and the higher the NO serum and C.S.F the more bad of prognosis (**table 12 and fig. 10**).

Tsukahara et al., (1998) reported that a higher proportion of patients with C.S.F nitrite above the normal range had neurologic or

audiologic sequelae (or both) in contrast to the patients with normal levels and this suggests that NO may be responsible for neurologic

damage in human.

In our study the children with septic meningitis were treated with intravenous ceftazidime (150 mg/kg in two divided doses) for 10 days and dexamethasone (0.6 mg/kg/day) for 4 days. The levels of NO in serum and C.S.F of those children were estimated before and after treatment. The result was significant decrease of NO levels in C.S.F and serum after treatment of those cases with antibiotics and dexamethasone.

These results are in agreement with *Jafari et al., (1994)* who reported that administration of dexamethasone with the commencement of antibiotic treatment is often recommended to improve the outcome of septic meningitis.

Kitteler et al., (1994) reported that corticosteroids are known to suppress NO synthase induction. Therefore, early dexamethasone therapy may be partly responsible for attenuation of increased NO production during recovery in patients with septic meningitis.

So we can conclude that, NO production is enhanced in the C.S.F and serum in cases of septic meningitis and support the hypothesis that NO contributes to the pathophysiology of septic meningitis. So NO can be used as simple and significant diagnostic and prognostic monitor of cases with septic meningitis but this needs further study in wider scale.

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